



ISSN : 0973-7057

Int. Database Index: 663 www.mjl.clarivate.com

Effect of intragastric daidzein administration on blood testosterone level in male Wistar rat.

Sweta Kumari* and Nayni Saxena

University Department of Zoology, Ranchi University, Ranchi, Jharkhand, India

Received : 12th July, 2019 ; Revised : 25th August, 2019

Abstract : Daidzein is a phytoestrogen found in many plants consumed by humans and animals. The influence of isoflavone, daidzein on blood testosterone hormone level was studied in sexually mature male Wistar rats. Rats were divided into three groups: Control (received no daidzein), Sham control [received vehicle as ethanol: saline water (1:9)] and Experimental group [received daidzein 15mg/kg body weight (bw)]. Daidzein was administered intragastrically, i.e. by using a cannula inserted via oesophagus into the stomach once a day for 30 continuous days. In animals treated with daidzein blood testosterone hormone level was significantly reduced as compared to the controls after 15 and 30 days. This effect was probably due to the direct inhibitory influence of daidzein on testosterone hormone secretion from testis. Results obtained in the study indicate that daidzein affects testosterone hormone secretion and may therefore be responsible for infertility, cryptorchidism and reduced sperm concentration in Wistar rat.

Key words: daidzein, testosterone, Wistar rat, isoflavone

INTRODUCTION

Daidzein (4',7, dihydroxyflavone) is a phytoestrogen belonging to class of soy isoflavones, a sub-class of flavonoids that have received great attention for their potential on human health benefits. Daidzein aglycone is an isoflavone found at low concentrations in soybeans but at high concentrations in certain soy-derived food. Daidzein is much more abundant in the unprocessed soybean. Some phytoestrogens have been categorized as endocrine disruptors, i.e. exogenous substances that alter the functions of the endocrine system and consequently have adverse effects on the health of the organism, its progeny or its populations. They mimic or block hormones and disrupt normal endocrine functions by altering normal hormone levels, inhibiting or stimulating the production of

hormones. They have been classified as natural environmental oestrogens¹ or natural endocrine- active agents². Structurally, daidzein closely resembles 17 β -estradiol and it binds to estrogen receptors (ERs), the stronger affinity being for the ER β isoform³. Acting as a natural selective ER modulator, daidzein exerts its estrogen agonist or antagonist action in tissue in a dose dependent manner⁴.

There is growing public concern regarding the adverse effects of environmental chemicals with an estrogenic influence on reproductive health. Daidzein and genistein are widely distributed in daily human diet⁵. In a typical Western diet, an average of 0.2 mg/kg isoflavones are consumed daily, whereas a typical Asian diet contains > 1.5 mg/kg isoflavones per day⁶, which can raise individual human isoflavone serum levels to 500 Nm⁷. For infants fed soy-based formulas isoflavone intake can reach 9.3 mg/kg bw/day.⁸ Phytoestrogens have been part of

*Corresponding author :

Phone : 7004792336

E-mail : swetatiwari3689@gmail.com

traditional diet in Asia for millennia. The negative effects of isoflavones, particularly on the male reproductive system have also been reported⁹. Previous studies demonstrated that isoflavones may produce male reproductive toxicity. Their main adverse effects on the male reproductive system include the disturbance of sex hormone release,^{10,11} interference with the onset of puberty¹², altering penile corpus cavernosum structure weakening erectile function^{13,14}, suppressing the activity of some steroidogenesis associated enzymes¹⁵ and decreasing the weight and epithelial height of accessory sex organs¹¹. A high intake of soy-based food and soy isoflavones is associated with reduced sperm concentration, as demonstrated in animal experiments and human epidemiological studies^{16,17}.

Dietary isoflavones have caused infertility in male mice¹⁸ and rats.¹⁹ Genistein and β - sitosterol had adverse effects on the reproductive organ weights in mice^{20,21} and rats.^{22,19} Reductions in the levels of plasma testosterone, estradiol and progesterone have been observed in rats after exposure to genistein^{23,21,24}. Phytoesterol exposure caused an increase in plasma testosterone levels in the European polecat²⁵ and in the field vole²⁶.

Keeping the above in mind and considering the fact that there are limited reports on daidzein, the present study was conducted to observe the effects of short term intragastric administration of the isoflavone, daidzein on testosterone hormone in male Wistar rat.

Experimental Design

The experiment was performed according to the guidelines accepted by the Ethics Committee, Ranchi University, Ranchi for Investigations on Animals.

Male and female Wistar rats weighing about 150g were used in the experiment. The animals were kept under standard conditions, at a constant temperature (21±4°C) with a 12-h dark-light cycle. Rats were fed a soy- free diet *ad libitum*. Males were divided into three groups of ten animals each. Two female rats were introduced in each group. Animals in the control group received no daidzein, whereas, rats in the second group received the vehicle, i.e. saline water: ethanol mixture (1:9 v/v). The third group (experimental) received daidzein dissolved in the vehicle in the amount of 15 mg/kg BW. The vehicle and daidzein solutions (Sigma) were administered intragastrically (0.5ml/150g BW) once a day for 30 consecutive days. The males were anesthetized (di-ethyl ether) and their blood serum were collected after 15 and 30 days of the start of the experiment. It was stored (-80°C) until analysis.

ANALYSIS

Testosterone hormone was determined by Fully Automated Bidirectional Interfaced Chemi Luminescent Immuno Assay.

RESULTS

Results for the effects of daidzein on serum testosterone level of rat after 15 and 30 days administration are presented in Table 1. & Figure 1.

Serum testosterone levels in animals administered daidzein intragastrically for all durations tested, were significantly (P<0.001) lower than their respective controls and sham-controls. No significant difference was observed between serum testosterone concentration of sham-operated animals and control animals

Table 1: Effect of intragastric administration of daidzein on serum testosterone level (ng/dl) of male Wistar rat

Treatment	15 days	30 days
Control	22.43 ± 0.08	22.7 ± 0.02
Sham control	22.29 ± 0.04	21.54 ± 0.08
Treated	7 ± 0.10*	7.5 ± 0.09*

Daidzein was dissolved in saline water: ethanol mixture (9:1 v/v) and was administered intragastrically (0.5 ml; 15 mg/kg BW) for 15days and 30 days.

Values are means ±SEM

Significance of difference from control:*, p<0.001.

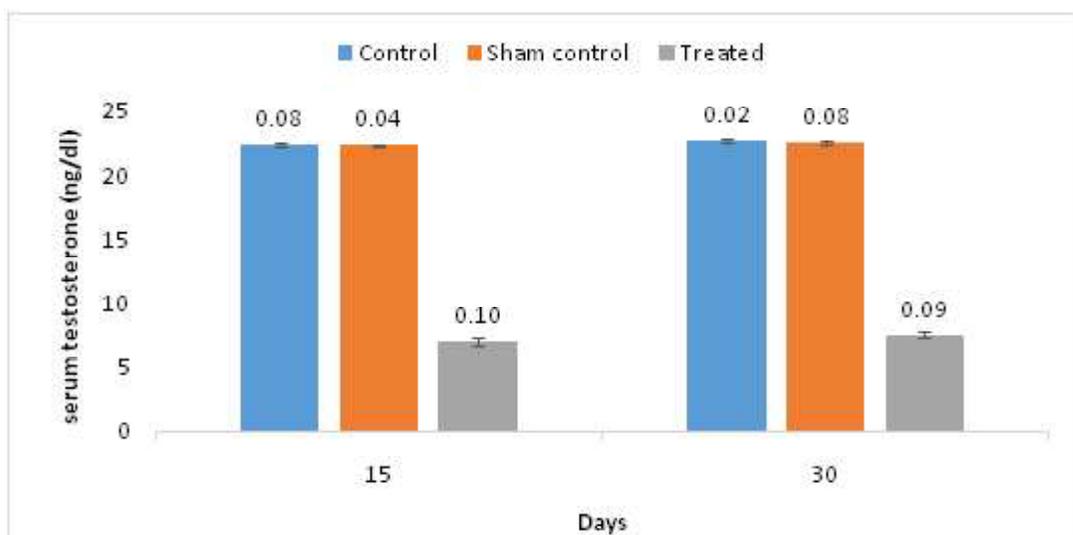


Figure 1: Effect of intragastric administration of daidzein (15mg/kg BW) for 15 days and 30 days on serum testosterone level (ng/dl) in male Wistar rat.

Significance of difference from control:*, p<0.001.

DISCUSSION

Studies for over the last 50 years reported an increased incidence of human male reproductive disorders like cryptorchidism, hypospadias, testicular cancer and low semen quality²⁷. The rapid increase in disorder of the reproductive function suggests the involvement of environmental or life style factors. Exposure to endocrine disruptors appears to be an important cause.

Endocrine disrupting chemicals are exogenous agents that interfere with synthesis, secretion, transport, metabolism, binding action or elimination of natural blood-borne hormones that are present in the body and are responsible for homeostasis, reproduction and developmental processes. These endocrine disruptors are highly heterogeneous in structure and widespread in our environment. Less attention has been paid to the action of natural plant-derived endocrine disruptors (phytoestrogens) which are non-steroidal compounds having the ability to bind to estrogen receptors.

Daidzein, the major soy isoflavone glucoside is present at high concentration in soybeans and is a major source of xenoestrogen exposure in humans and animals. The results of the present study indicate an inhibitory effect of daidzein on the testosterone hormone level in the serum of Wistar rat when administered at a dose of 15 mg/kg body weight for a period of 15 days. Study on the long

term administration of daidzein, at this dose, for over 30 days showed the same inhibitory action of daidzein on this hormone. The present result clearly shows that this isoflavone i.e. daidzein, under the present experimental conditions, is able to alter the testosterone level in the blood serum of this animal. This could have potentially detrimental effects on the fertility and reproductive functions of the animal.

Adverse effects of phytoestrogens have been observed in experimental animals but almost all of them focused on the effects of genistein and less attention has been paid to study the action of daidzein on reproductive functions. For example, the adverse effects of phytoestrogen, genistein in causing infertility in male mice were observed long back¹⁸. It has been shown that exposure to genistein adversely affects the reproductive organ weights in mice^{20,21} and rats^{19,22}.

Reduction in the plasma testosterone, estradiol and progesterone levels, after genistein exposure have been shown in rats^{21,23}. However, such reports are lacking for daidzein.

The present results are new and show that the isoflavone daidzein is capable of causing an effect on the testosterone level in the plasma, of this animal. These results are different from those observed for the European polecat, *Mustela putorius*,²⁶ in which it was demonstrated that

phytoesterol exposure increased the plasma testosterone levels.

Studies have reported that certain foods containing isoflavones, particularly infant formula, may have potential adverse effects on male reproductive function. Humans and animals are frequently exposed to products that contain low levels of isoflavones²⁸. There are commercially available isoflavone containing products that have estrogen – like effects such as milk with added genistein and soy-based infant formulas^{29,30}.

Daidzein is found in many natural products that are marketed as promoting health and used to treat various diseases such as asthma, cancer, menopause, osteoporosis, atherosclerosis³¹ as well as many commercial pet foods³² and soy based infant formulas^{33,29}.

Isoflavones cause infertility in many animal species like sheep³⁴, the California quail³⁵, the mouse^{36,18}, the cheetah³⁷ and the rat¹⁹. Dietary phytoestrogens may have similar effects on the development and fertility of other species, including humans. But human data are very limited. There is paucity of studies available to evaluate the effects of soy and phytoestrogens on fertility or reproductive hormone in human males. Associations between exposure to phytoestrogens and hormonal disorders in children have been described³⁰.

The present results clearly demonstrated that dietary daidzein administration was able to decrease testosterone synthesis just after a period of 15 days in rats. These results are similar to those obtained by Zhu and Coworkers³⁸ who reported a suppression of testosterone secretion by daidzein (30 μ mol /l for 24 hrs) in cultured testes and Leydig cells of neonatal mouse testes. These authors also suggested adverse effects of high doses of daidzein on Sertoli cells in neonatal mouse testes. A similar effect for genistein in impairing early testosterone production in fetal mouse testes *in vitro* has also been reported³⁹.

Genistein at a dose of 213 mg/kg bw/day from gestational day 7 to PND 13 also reduced plasma testosterone levels in rat offspring⁴⁰. A reduction in plasma testosterone level was detected in primary Leydig cells by daidzein treatment in *Anser anser*^{41,42}. The findings show a potential harmful effect of daidzein exposure on testis steroidogenesis function during the early neonatal period.

The results of the present study clearly define the association between daidzein exposure and reduced

testosterone production in Wistar rat, under the tested experimental conditions. The mechanism of this action is still unclear. It has been suggested that testosterone production can be inhibited by exogenous compounds by suppression of steroidogenic enzymes, P450 scc, 3 β -HSD and P450c17 β ⁴³⁻⁴⁶. It has been reported that genistein suppresses StAR, P450 scc, 3 β -HSD and P450C17 β expression and decreases testosterone production in fetal testis³⁹. The observations in these studies may explain why testosterone levels in the testis or plasma are decreased. The expression levels of STAR and steroidogenic enzymes have been evaluated in daidzein- treated Leydig cells, in order to verify the mechanistic activities of daidzein on factors associated with steroid synthesis. The results showed decline in m-RNA and protein expression levels of STAR, P450scc, 3 β - HSD and P450C17. These results have been reported to be consistent with those of organ culture⁴⁷. Testosterone is synthesized from cholesterol.

The conversion of cholesterol to testosterone involves series of key steroidogenic proteins, such as STAR protein, cytochrome p450 scc and 3 β -HSD^{48,49,50}. The rate limiting step in the synthesis of testosterone is the transfer of cholesterol to pregnenolone via P450scc; StAR mediates this rate-limiting step in steroidogenesis⁵¹. The conversion of pregnenolone to progesterone is mediated by 3 β -HSD. The enzymatic action is essential for the production of all active steroid hormones⁵².

Phytoestrogens including daidzein can modulate the endocrine system by altering the gene expression or activity of the enzymes involved in steroidogenesis⁵³. The key enzymes involved in steroidogenesis have been reported to be important targets for phytoestrogens⁵⁴. Isoflavones including daidzein can exert inhibitory feedback on the hypothalamus-pituitary gonadal axis⁵⁵.

The present results confirm the inhibitory effect of dietary daidzein on testosterone levels of blood in Wistar rat. It is suggested that this effect of daidzein may be exerted via the alteration of the activity of the enzymes involved in steroidogenesis. Further research in this direction will elucidate the complete mechanism involved in producing such an effect.

CONCLUSION

The results obtained in this study prove that 15 and 30 days intragastric administration of daidzein affects

testosterone hormone level in male Wistar rats. This compound essentially decreased testosterone concentration in blood serum. Mechanisms responsible for the detected activity of daidzein require further investigations.

REFERENCE

1. Arukwe, A. 2001. Cellular and molecular responses to endocrine-modulators and the impact on fish reproduction. *Mar Pollut Bull.* **42**: 643-655.
2. Safe, K., Connor, K., Ramamoorthy, K., Gaido, K., Maness, S. 1997. Human exposure to endocrine active chemicals: hazard assessment problems. *Regul Toxicol Pharmacol.* **26**:52-58.
3. Kuiper, G.G.J.M., Lemmen, J.G., Carsson, B., Corton, C. J., Safe, S.H., van der Saag, P.T., van der Burg, B., Gustafsson, J.-Å. 1998. Interaction of estrogenic chemicals and phytoestrogens with estrogen receptor. *Endocrinology.* **139**: 4252-4263.
4. Setchell K. D. 2001. Soy isoflavones-benefits and risks from nature's selective estrogen receptor modulators (SERMs). *J. Am. Coll. Nutr.* **20**: 354S-362S.
5. Kurzer M.S., Xu X. 1997. Dietary phytoestrogens. *Annu. Rev. Nutr.* **17**: 353-381.
6. Coward, L., Barnes, N.C., Setchell, K.D.R., Barnes, S. 1993. Genistein, daidzein, and their β -glycoside conjugates: antitumor isoflavones in soybean foods from American and Asian diets. *J Agric Food Chem.* **41**:1961-1967.
7. Morton MS, Arisaka O, Miyake N, Morgan LD and Evans BA. 2002. Phytoestrogen concentrations in serum from Japanese men and women over forty years of age. *J Nutr.* **132**: 3168-3171.
8. McCarver G, Bhatia J, Chambers C, Clarke R, Etzel R, Foster W, Hoyer P, Leeder JS, Peters JM, Rissman E. 2011. NTPCERHR expert panel report on the developmental toxicity of soy infant formula. *Birth Defects Res B Dev. Reprod. Toxicol.* **92**: 421-468.
9. Rozman KK, Bhatia J, Calafat AM, Chambers C, Culty M, Etzel RA, Flaws JA, Hansen DK, Hoyer PB, Jeffery EH. 2006. NTP CERHR expert panel report on the reproductive and developmental toxicity of genistein. *Birth Defects Res. B Dev Reprod Toxicol.* **77**: 485-638.
10. Akingbemi BT, Braden TD, Kemppainen BW, Hancock KD, Sherrill JD, Cook SJ, He X and Supko JG 2007. Exposure to phytoestrogens in the perinatal period affects androgen secretion by testicular Leydig cells in the adult rat. *Endocrinology.* **148**: 4475-4488.
11. Yuan XX, Zhang B, Li LL, Xiao CW, Fan JX, Geng MM and Yin YL 2012. Effects of soybean isoflavones on reproductive parameters in Chinese mini pig boars. *J Anim Sci Biotechnol.* **3**: 31.
12. Caceres S, Pena L, Moyano G, Martinez Fernandez L, Monsalve B, Illera MJ, Millan P, Illera JC and Silvan G. 2015. Isoflavones and their effects on the onset of puberty in male Wistar rats. *Andrologia.* **47**: 1139-1146.
13. Huang Y., Pan L., Xia X., Feng Y., Jiang C. and Cui Y. 2008. Long term effects of phytoestrogen daidzein on penile cavernosal structures in adult rats. *Urology.* **72**: 220-224.
14. Pan L, Xia X, Feng Y, Jiang C and Huang Y. 2007. Exposure to the phytoestrogen daidzein attenuates apomorphine induced penile erection concomitant with plasma testosterone level reduction in dose and time related manner in adult rats. *Urology.* **70**: 613-617.
15. Hu GX, Zhao BH, Chu YH, Zhou HY, Akingbemi BT, Zheng ZQ and Ge RS. 2010. Effects of genistein and equol on human and rat testicular 3 β hydroxysteroid dehydrogenase and 17 β hydroxysteroid dehydrogenase 3 activities. *Asian J Androl.* **12**: 519-526.
16. Chavarro JE, Toth TL, Sadio SM and Hauser R 2008. Soy food and isoflavone intake in relation to semen quality parameters among men from an infertility clinic. *Hum Reprod.* **23**: 2584-2590.
17. Cederroth CR, Zimmermann C, Beny JL, Schaad O, Combepine C, Descombes P, Doerge DR, Pralong FP, Vassalli JD and Nef S. 2010. Potential detrimental effects of a phytoestrogen rich diet on

- male fertility in mice. *Mol. Cell Endocrinol.* **321**: 152-160.
18. **East, J. 1955.** The effect of genistein on the fertility of mice. *J Endocrinol.* **13**: 94-100.
19. **Nagao, T., Yoshimura, S., Saito, Y., Nakagomi, M., Usumi, K., Ono, H. 2001.** Reproductive effects in male and female rats of neonatal exposure to genistein. *Reprod. Toxicol.* **15**: 399-411.
20. **Strauss, L., Mäkelä, S., Joshi, S., Huhtaniemi, I., Santti, R. 1998.** Genistein exerts estrogen-like effects in male mouse reproductive tract. *Mol Cell. Endocrinol.* **144**: 83-93.
21. **Wisniewski, A.B., Cernetich, A., Gearhart, J.P., Klein, S.L. 2005.** Perinatal exposure to genistein alters reproductive development and aggressive behavior in male mice. *Physiol. Behav.* **84**: 327-334.
22. **Malini, T. and Vanitha Kumari, G. 1991.** Antifertility effects of β -sitosterol in male albino rats. *J. Ethnopharmacol.* **35**:149-153.
23. **Awad, A.B., Hartati, M.S., Fink, C.S. 1998.** Phytosterol feeding induces alteration in testosterone metabolism in rat tissues. *J. Nutr. Biochem.* **9**: 712-717.
24. **Awoniyi, C.A., Roberts, D., Veeramachaneni, R., Hurst, B.S., Tucker, K.E., Schlaff, W.D. 1998.** Reproductive sequelae in female rats after in utero and neonatal exposure to the phytoestrogen genistein. *Fertil Steril.* **70**: 440-447. Nieminen, P., Mustonen, A.-M., Lindström-Seppä, P.,
25. **Asikainen, J., Mussalo-Rauhamaa, H., Kukkonen, J.V.K. 2003b.** Phytosterols act as endocrine and metabolic disruptors in the European polecat (*Mustela putorius*). *Toxicol. Appl. Pharmacol.* **178**: 22-28.
26. **Nieminen, P., Mustonen, A.-M., Lindström-Seppä, P., Kärkkäinen, V., Mussalo-Rauhamaa, H., Kukkonen, J.V.K. 2003c.** Phytosterols affect endocrinology and metabolism of the field vole (*Microtus agrestis*). *Exp. Biol. Med.* **228**:188-193.
27. **Skakkeback et al., 2001.**
28. **Eustache F, Mondon F, Canivenc-Lavier MC, Lesaffre C, Fulla Y, Berges R, Cravedi JP, Vaiman D, Auger J. 2009.** Chronic dietary exposure to a low-dose mixture of genistein and vinclozolin modifies the reproductive axis, testis transcriptome, and fertility. *Environ Health Perspect.* **117**:1272–1279.
29. **Setchell KD, Zimmer-Nechmias L, Cai J, Heubi JE. 1998.** Isoflavone content of infant formulas and the metabolic fate of these phytoestrogens in early life. *Am J Clin. Nutr.* **68**:1453–1461.
30. **Greim HA 2004.** The endocrine and reproductive system: adverse effects of hormonally active substances? *Pediatrics.* **113**:1070-1075.
31. **Setchell, K.D.R., Brown, N.M., Desai, P., Zimmer-Nechmias, L., Wolfe, B.E., Brashear, W.T., Kirschner, A.S., Cassidy, A., Heubi, J.E. 2001.** Bioavailability of pure isoflavones in healthy humans and analysis of commercial soy isoflavone supplements. *J Nutr.* **131**: 1362S-1375S.
32. **Brown, N.M. and Setchell, K.D.R. 2001.** Animal models impacted by phytoestrogens in commercial chow: implications for pathways influenced by hormones. *Lab Invest.* **81**: 735-747.
33. **Setchell, K.D.R., Zimmer-Nechmias, L., Cai, J., Heubi, J.E. 1997.** Exposure of infants to phytoestrogens from soy-based infant formula. *Lancet.* **350**: 23-27.
34. **Bennetts, H.W., Underwood, E.J., Shier, F.L.A. 1946.** A specific breeding problem of sheep on subterranean clover pastures in Western Australia. *Aust J. Agric. Res.* **22**: 131-138.
35. **Leopold, S.A., Erwin, M., Oh, J., Browning, B. 1976.** Phytoestrogens: adverse effects on reproduction in California quail. *Science.* **191**: 98-100.
36. **Jefferson, W.N., Padilla-Banks, E., Newbold, R.R. 2005.** Adverse effects on female development and reproduction in CD-1 mice following neonatal exposure to the phytoestrogen genistein at environmentally relevant doses. *Biol Reprod.* **73**:798-806.

37. Setchell, K.D., Gosselin, S.J., Welsh, M.B., Johnston, J.O., Balisteri, W.F., Kramer, L.W., Dresser, B.L., Tarr, M.J. 1987. Dietary estrogens- a probable cause of infertility and liver disease in captive cheetahs. *Gastroenterology*. **93**: 225-233.
38. Zhu Y., Xu H., Li M., Gao Z., Huang J., Liu L., Huang X., Li Y. 2016. Daidzein impairs leydig cell testosterone production and sertoli cell function in neonatal mouse testes: an *in vitro* study. **315**: 5325-5333.
39. Lehraiki A, Chamaillard C, Krust A, Habert R and Levacher C. 2011. Genistein impairs early testosterone production in fetal mouse testis via estrogen receptor alpha. *Toxicol In Vitro* **25**: 1542-1547.
40. Boberg J, Mandrup KR, Jacobsen PR, Isling LK, Hadrup N, Berthelsen L, Elleby A, Kiersgaard M, Vinggaard AM, Hass U and Nellemann C. 2013. Endocrine disrupting effects in rats perinatally exposed to a dietary relevant mixture of phytoestrogens. *Reprod Toxicol*. **40**: 41-51.
41. Opalka DM, Kaminska B, Piskula MK, Puchajda Skowronska H and Dusza L. 2006. Effects of phytoestrogens on testosterone secretion by Leydig cells from Bilgoraj ganders (Anseranser). *Br Poult Sci*. **47**: 237-245.
42. Opalka M, Kaminska B, Leska A and Dusza L. 2012. Mechanism of phytoestrogen action in Leydig cells of ganders (Anseranser domesticus): Interaction with estrogen receptors and steroidogenic enzymes. *J. Environ. Sci. Health A. Tox. Hazard Subst. Environ. Eng*. **47**:1335-1339.
43. Hannas BR, Lambright CS, Furr J, Evans N, Foster PM, Gray EL and Wilson VS. 2012. Genomic biomarkers of phthalate induced male reproductive developmental toxicity: A targeted RT-PCR array approach for defining relative potency. *Toxicol Sci*. **125**: 544-557.
44. Ye L, Su ZJ and Ge RS 2011. Inhibitors of testosterone biosynthetic and metabolic activation enzymes. *Molecules*. **16**: 9983-10001.
45. Liu S, Wang D, Zhang J, Zhang D, Gong M, Wang C, Wei N, Liu W, Wang Y, Zhao C, 2012. Citrinin reduces testosterone secretion by inducing apoptosis in rat Leydig cells. *Toxicol In Vitro*. **26**: 856-861.
46. N'Tumba Byn T, Moison D, Lacroix M, Lecureuil C, Lesage L, Prud homme SM, Pozzi Gaudin S, Frydman R, Benachi A, Livera G. 2012. Differential effects of bisphenol A and diethylstilbestrol on human, rat and mouse fetal Leydig cell function. *PLoS One*. **7**:551-579.
47. Delbès G, Duquenne C, Szenker J, Taccoen J, Habert R and Levacher C. 2007. Developmental changes in testicular sensitivity to estrogens throughout fetal and neonatal life. *Toxicol Sci*. **99**: 234-243.
48. Cherradi N, Rossier MF, Vallotton MB, Timberg R, Friedberg I, Orly J. 1997. Submitochondrial distribution of three key steroidogenic proteins (steroidogenic acute regulatory protein and cytochrome P450_{scc} and 3 α -hydroxysteroid dehydrogenase isomerase enzymes) upon stimulation by intracellular calcium in adrenal glomerulosa cells. *J Biol Chem*. **272**: 7899-907.
49. Stocco DM 1998. Recent advances in the role of StAR. *Rev Reprod*. **3**: 82-5.
50. Walsh LP, Webster DR, Stocco DM. 2000. Dimethoate inhibits steroidogenesis by disrupting transcription of the steroidogenic acute regulatory (StAR) gene. *J. Endocrinol*. **167**: 253-63.
51. Stocco DM, Clark BJ. 1996. Regulation of the acute production of steroids in steroidogenic cells. *Endocr Rev*. **17**: 221-44.
52. Payne AH, Hales DB. 2004. Overview of steroidogenic enzymes in the pathway from cholesterol to active steroid hormones. *Endocr Rev*. **25**: 947-70.
53. Hilscherova K, Jones PD, Gracia T, Newsted JL, Zhang X, Sanderson JT. 2004. Assessment of the effects of chemicals on the expression of ten

Biospectra : Vol. 14(2), September, 2019

An International Biannual Refereed Journal of Life Sciences

- steroidogenic genes in the H295R cell line using real-time PCR. *Toxicol Sci.* **81**:78-89.
- 54. Sanderson JT. 2006.** The steroid hormone biosynthesis pathway as a target for endocrine-disrupting chemicals. *Toxicol Sci.* **94**:3-21.
- 55. Kwon SM, Kim SI, Chun DC, Cho NH, Chung BC, Park BW. 2001.** Development of rat prostatitis model by oral administration of isoflavone and its characteristics. *Yonsei Med J.* **42**:395-404.
